

into the reaction mixtures to give **3,3'a-e** quantitatively. Because **1** has a five-membered ring, reaction to form a spirobicycle system was favored.

Pathway B: under a nitrogen atmosphere at 25 °C 1 equiv. of phenol in benzene or dichloromethane was added dropwise to a benzene or dichloromethane solution of 2,2,2-trichloro-1,3,2-benzodioxaphosphole and stirred continuously. A few minutes later, 2,2-dichloro-2-phenoxy-1,3,2-benzodioxaphosphole **4** was quantitatively obtained, showing one peak in the ³¹P NMR spectrum at δ -34.13. Then an *N,O*-bis(trimethylsilyl)amino acid **a-e** was added to the reaction mixture to give **3,3'a-e**. Compounds **3,3'a-d** were crystallized from dichloromethane-diethyl ether (1:5) while **3,3'e** was recrystallized from benzene, to give the products, in some cases, as white solids; yields: 63–71% (pathway A); 74–83% (pathway B).

Since the P–O bond of the anhydride P–O–CO moiety was longer than expected in the pentacoordinate phosphorane,¹⁶ it implies that this is the apical bond. Indeed, each of the compounds **3,3'a-e** had a large ²*J*_{P–OCO}-value. Because glycine has no chiral carbon, **2a,2'a** only showed one peak in the ³¹P NMR spectrum at δ _p -27.89, and their derivatives **3a,3'a** also showed only one peak at δ _p -42.02. The anhydride POC=O bond is of high energy,¹⁷ and consequently yields a peptide bond when the amino group of an amino acid attacks the carboxy group of the anhydride. Hence, the pentacoordinate imino(alkyl)acetoxyposphoranes might be intermediates in protein biosynthesis and could have potential in polypeptide synthesis. In summary, the high yields, rapid reactions and easy availability of the starting materials and reagents render this synthesis an efficient process.

Experimental

¹H, ¹³C, ¹H–¹H COSY and ¹H–¹³C COSY NMR spectra were recorded on a Bruker AM-500 spectrometer at 500 MHz in CDCl₃ solvent with chemical shifts referenced to CDCl₃ (δ _H = 7.24, δ _C = 77). *J* Values are given in Hz. ³¹P NMR spectra were determined by a Bruker AM-200 spectrometer at 200 MHz using 85% H₃PO₄ (δ _p = 0) as an external standard. Elemental analyses were carried out on a Carlo Erba 1106 CHN analyzer. Field desorption mass spectra were obtained on a Finnigan MAT 90 double-focusing mass spectrometer.

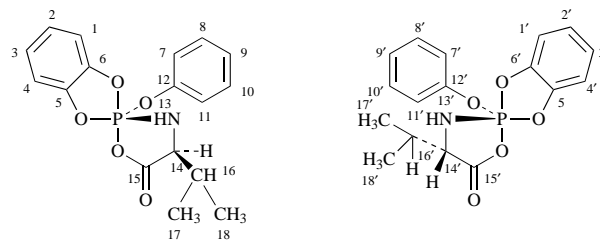
The preparation of compound **3,3'c** is given as an example.

Pathway A

To a stirred solution of **1** (1.23 g, 5 mmol) in anhydrous benzene (15 ml) at room temperature under a nitrogen atmosphere was added dropwise *N,O*-bis(trimethylsilyl)valine (1.31 g, 5 mmol) in benzene (10 ml). After 10 min, 0.5 ml of the mixture was withdrawn and its ³¹P NMR spectrum was taken (δ _p **1**: -26.23; **2c,2'c**: -29.39, -29.92). Phenol (0.47 g, 5 mmol) in benzene (10 ml) was then added dropwise to the mixture. After 10 min the precipitate was filtered off, the solvent and trimethylchlorosilane were removed by rotary evaporation, and the remaining solution (ca. 1 ml), was diluted with dry diethyl ether (5 ml). A white precipitate appeared which was filtered off, and the filtrate was washed three times with dry diethyl ether and dried *in vacuo* over P₂O₅ at 39 °C for 5 h to afford **3,3'c** (2.32 g, 67%).

Pathway B

To a stirred solution of 2,2,2-trichloro-1,3,2-benzodioxaphosphole (1.23 g, 5 mmol) in anhydrous benzene (15 ml) at room temperature under a nitrogen atmosphere was added dropwise phenol (0.47 g, 5 mmol) in benzene (10 ml). After 10 min, 0.5 ml of the mixture was withdrawn and its ³¹P NMR spectrum was taken (δ _p **1**: -26.23; **4**: -34.13). *N,O*-Bis(trimethylsilyl)valine (1.31 g, 5 mmol) in benzene (10 ml) was then added dropwise to the mixture. After 10 min the reaction was worked-up as for Pathway A. Isolated yields and purities were similar to pathway



A. Yield of **3,3'c**: 2.56 g, 74%; mp 148–150 °C (Found: C, 58.75; H, 5.18; N, 4.05. C₁₇H₁₈NO₅P requires C, 58.79; H, 5.19; N, 4.03%); δ _H 0.92 (d, 3 H, 17-CH₃), 0.99 (d, 3 H, 18-CH₃), 1.10 (m, 6 H, 17'- and 18'-CH₃), 2.18 (m, 1 H, 16-CH), 2.26 (m, 1 H, 16'-H), 3.76 (d, 1 H, 13-NH, ²*J*_{PNH} 16.19), 3.81 (d, 1 H, 13'-NH, ²*J*_{PNH} 16.19), 3.87 (m, 1 H, 14-CH), 3.88 (m, 1 H, 14'-CH), 6.56–6.68 (dd, 2 H, 1,1'-Ph), 6.81–6.94 (m, 8 H, 2,2',3,3'-Ph, 7,7',11,11'-Ph), 7.07–7.13 (m, 4,4'-Ph, 9,9'-Ph) and 7.21–7.23 (m, 8,8',10,10'-Ph); δ _C 16.21 (17-CH₃), 16.86 (17'-CH₃), 18.61 (18-CH₃), 19.15 (18'-CH₃), 31.04 (d, 16-CH), 31.12 (d, 16'-CH), 60.45 (14-CH), 60.54 (14'-CH), 109.92 (d, 1-Ph), 110.08 (d, 1'-Ph), 111.47 (d, 4-Ph), 111.60 (d, 4'-Ph), 120.78 (3,3',7,7',11,11'-Ph), 123.51 (2,2'-Ph), 124.89, 125.10 (9,9'-Ph), 129.26, 129.47 (8,8',10,10'-Ph); 142.22 (5,5'-Ph), 144.96 (6,6'-Ph), 152.06 (12,12'-Ph), 167.99, 168.17 (dd, 15,15'-CO-, ²*J*_{P-C} 17.95); δ _p -44.64 and -44.83; *m/z* (FDMS) 347 (M⁺).

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References

- (a) F. Ramirez, *Acc. Chem. Res.*, 1968, **1**, 168; (b) I. Ugi, D. Marquarding, H. Klusacek, P. Gillespie and F. Ramirez, *ibid*, 1971, **4**, 288.
- (a) S. J. Benkovic and K. J. Schray, *Acc. Chem. Res.*, 1969, **91**, 5653; (b) G. D. Smith, C. N. Caughlan, F. Ramirez, S. L. Glaser and P. Stern, *ibid*, 1974, **96**, 2698.
- V. M. Clark and A. J. Kirby, *J. Am. Chem. Soc.*, 1963, **85**, 3705.
- (a) S. J. Benkovic and K. J. Schray, *Biochemistry*, 1968, **7**, 4090; (b) K. J. Schray and S. J. Benkovic, *J. Am. Chem. Soc.*, 1971, **93**, 2522.
- G. J. Ji, C. B. Xue, J. N. Zeng, L. P. Li, W. G. Chai and U. F. Zhao, *Synthesis*, 1988, 444.
- J. N. Zeng, C. B. Xue, Q. W. Chen and Y. F. Zhao, *Bioorg. Chem.*, 1989, **17**, 434.
- C. B. Xue, J. Z. Wu, Y. W. Yin and Y. F. Zhao, *J. Chem. Soc., Perkin Trans. 2*, 1990, **2**, 431.
- X. B. Ma and Y. F. Zhao, *J. Org. Chem.*, 1989, **54**, 4005.
- D. Q. Zhang, Y. F. Zhao and C. B. Xue, *Int. J. Pept. Protein Res.*, 1991, **39**, 457.
- Y. F. Zhao, Y. Ju, Y. M. Li, Q. Wang, Y. W. Yin and B. Tan, *Int. J. Pept. Protein Res.*, 1995, **45**, 514.
- W. H. Zhou, Y. Ju, Y. F. Zhao, Q. G. Wang and G. A. Luo, *Origins Life Evol. Biosphere*, 1996, **26**, 547.
- Q. Wang, Y. F. Zhao, F. L. An, Y. Q. Mao and J. Z. Wang, *Sci. China, Ser. B*, 1995, **25**, 684.
- F. Lipmann, *Adv. Enzymol. Relat. Subj. Biochem.*, 1941, **1**, 97.
- E. D. Smith and H. Sheppard, *Nature*, 1965, **208**, 878.
- T. A. Khwaja, C. B. Reese and J. C. M. Stewart, *J. Chem. Soc. C*: 1970, 2092.
- G. D. Smith, C. N. Caughlan, F. Ramirez, S. L. Glase and P. Stern, *J. Am. Chem. Soc.*, 1974, **96**, 2698.
- D. M. Hayes, G. L. Kenyon and P. A. Kollman, *J. Am. Chem. Soc.*, 1975, **97**, 4762.
- H. Fu, Z. L. Li and Y. F. Zhao, submitted for publication in *J. Am. Chem. Soc.*

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